Celvol (Polyvinvyl Chloride) 205 (805 used only if 205 is not available)

(This is a hardening aqueous mounting medium for use with samples in PBS like tissue sections stained with X-gal or HRP/DAB without dehydration. It is not compatible with tissue sections that have been through organic solvents like xylene.)

50 g Celvol
160 mL PBS (without azide)
80 mL Glycerol

1) Add to room temperature PBS while stirring (use largest stirring bar possible)
2) Set temperature setting between 3-4
3) After stirring, bring temperature to 90˚C setting (watch that does not boil, since once it boils, the solution burns and the solution will have a slight brownish tint to it)—the temperature does not need to be precisely 90˚C (needs to be very warm, celvol gets added slowly)
4) Cook for 30 minutes or until it is a mass of mini-bubbles (if stir bar not moving, then use a spatula to scrape the bottom of the beaker)
5) Let cool until warm to touch or wait 15 minutes (do not let it cool completely)
6) Skim off any mass floating at the top
7) Stir in glycerol manually with a disposable pipet
8) Transfer to 15 mL Falcon tubes
9) Spin at 1500 rpm for 5 minutes (gets rid of bubbles)

USE: Store in the 15 ml tubes for years at room temperature. It is gooey; try not to introduce bubbles, but they can always be centrifuged away. If too viscous for the disposable pipette, cut off some of the pipette's tip. However, you may find that enough coats the outside of the pipette that you don't even need to fill it. Keep the cap closed as much as possible; otherwise the tube will start drying out. Mount coverslips and let dry overnight to solidify; although slides can be viewed immediately if the coverslips are not disturbed.

Supplier: Celanese Ltd.
P.O.Box 819005
Dallas, TX 75381
972-443-4000

celvol V-205
Polyvinyl Alcohol
supplied as free sample, enough to last for several years for this application

Matt Springer, updated 3/3/06 from a Blau lab protocol