

## Hematoxylin & Eosin Staining of Sections

(If only eosin staining, just follow pink directions)

Cut frozen sections at 10 $\mu$ m onto gelatin-coated slides

Air dry

Stain immediately or store at -20 $^{\circ}$

Filter Hematoxylin before use to remove gold film

(or can skim the top of the hematoxylin in the staining dish with a kimwipe; wear a lab coat!!)

Hematoxylin - 3 min (Harleco)

Wash 2X tap water

Blue in Scotts tap water - 1 min (\*\*)

Wash tap water

Eosin - 30 s (Eosin Y, Sigma)

95% EtOH - 1 min

95% EtOH "

100% EtOH "

100% EtOH "

50/50 EtOH/Xylene - 1 min

Xylene - 1 min

Xylene "

Mount with organic mounting medium like Pro-Texx or Permount

NOTES: You can re-use the eosin for a while but don't add it back in to unused eosin, because used eosin + new eosin = used eosin.

During dehydration series, let the previous liquid drain off for a few seconds (but don't let sections dry) to transfer as little as possible into the next liquid.

(\*\*)Scotts tap water

2.0 gm Sodium Bicarbonate

20.0 gm Magnesium Sulfate

1 L tap water

can add a pinch of thymol to retard mold